

Quorum: 7

IBC Meeting Comments

February 18, 2026 – Zoom Conference Call

Attendance – IBC Committee			
Present	Name	Expertise	Role
	Paul Gulig	Microbiology	Member
	Luis Martinez	Microbiology	Member
x	Jason Clements		Member
	Norman Beatty	Infectious Diseases	Member
x	Steeve Boulant Joined at 1:35pm	Virology	Member
x	Sanford L. Boye	Virology	Member
	Amber Duren		Community Member
x	Mariola Ferraro	Microbiology	Member
x	Gary Heil		Member
x	Michael McIntosh	Virology	Member
x	John D. McVay	Plants	Community Member
	Mark Moehle	Microbiology	Member
x	Kamal A. Mohammed	Microbiology	Member
x	Christopher Overend		Member
x	[REDACTED]	Animals	Member
x	Jeffrey Rollins Joined at 1:23pm	Plants	Member
x	Elias J. Sayour Joined at 2:02pm	Clinical Trials	Member
x	Clay Smith	Virology	Chair
	Daniel R. Swale	Insects	Member
	Amy Vittor	Infectious Diseases	Member

Attendance – Staff and Guests		
Present	Name	Affiliation/Position
x	Asha Rani	UF EHS
x	Christine Lashley	UF EHS
x	Pratibha Srivastava	UF EHS
x	Alek Arnayos	UF EHS
x	[REDACTED]	UF IACUC
x	Laura Castillo	UF EHS
x	Jennifer Jackson	UF EHS
x	Erica Gonzaga	UF EHS
x	Anna Gioseffi	UF EHS
x	Raies Mir	UF EHS
x	Laurence Prunetti Joined at 1:04pm	UF EHS
x	Savannah Hardiman Joined at 1:09pm	UF EHS
x	Artiom Chacon Joined at 1:15pm	UF EHS
x	Larry Burris	IBC Guest

Attendance – UFO IBC: Apopka, Orlando, Lake Alfred, Lake Nona		
Present	Name	Affiliation/Position
	Annette R. Khaled	Community member - Covers Apopka, Orlando, Lake Alfred and Lake Nona
	Norman Beatty	Other: Infectious Disease
x	Mariola J. Ferraro	Other: Microbiology and Cell Science
x	Gary L. Heil	Other: Biosafety/Alternate Responsible Official
	Kindra A. Kelly-Quagliana	BSO, Voting Contact
x	Michael T. McIntosh	Other: infectious disease
x	Harvey E. Ramirez	Animal Expert
x	Jeffrey Rollins	Plant Expert
	Hubert Salvail	Community member - Covers Apopka, Orlando, Lake Alfred and Lake Nona
x	Wesley Clay Smith	Chair

Agenda:

	Full Committee Projects	PI
1	Investigating mechanisms of brain function in health and stress-induced disease	██████████ BIO5709 Renewal
2	Sea anemone in situ hybridization	██████████
3	Induction of T-Cell Tolerance to an auto-antigen by AAV hepatic gene therapy	██████████ BIO5687 Renewal
4	Functional characterization of genes conferring virulence at low temperatures in <i>Ralstonia solanacearum</i> Will be voted on by UF Orlando IBC	David Norman RD-3522 Transcription

5	Detection of nucleic acids using CRISPR	Piyush Jain BIO5522 Renewal
6	Engineering immune cells for cancer immunotherapy	██████████ BIO5642 Renewal
7	A Phase 1 Study of FT819 in B-cell Mediated Autoimmune Diseases	Michael Bubb
8	Neuromodulation of smell and reward	Karina Alvina BIO5764 Renewal
Amendments		PI
1	Post-transcriptional regulation of gene expression in neuromuscular disease	██████████ BIO5876
2	Research on Clavibacter species, wilt pathogens of maize, tomato, alfalfa and potato	Frank White – BIO6122
3	Mature Citrus Transformation and Gene Editing Service Will be voted on by UF Orlando IBC	Janice Zale – BIO6099
4	A Phase 1 Study of Inhaled KB4808 for the Treatment of Alpha-1 Antitrypsin Deficiency	Jorge Lascano – BIO7138
5	The role of glucan phosphatases in metabolism	██████████ – BIO6425
6	Use of rodent models of disease to develop biomarkers and therapeutics	██████████ – BIO6129

Minutes

Meeting was called to order at: 1:02pm

Project Review

Principal Investigator: [REDACTED] BIO5709 Renewal

Project Title: Investigating mechanisms of brain function in health and stress-induced disease

BIO5709

Vector/Agent(s) to be used: Adeno-associated virus (AAV)

Name and Function of Transcribed Nucleic Acids: RNA, increase brain activity and promote neuroprotection, increase cellular metabolism, silencing cells expressing NPY (neuropeptide Y), protein with metabolic functions. RNA to target Interleukin-6, IL-6 is a mediator of neuroinflammation, and is involved in neurodegeneration and unhealthy aging; thus, using a silencing AAV will reduce the levels of the cytokine and it is expected that this will result in neuroprotection. RNA to target TNF-alpha, TNF-alpha is a mediator of neuroinflammation, and is involved in neurodegeneration and unhealthy aging; thus, using a silencing AAV will reduce the levels of the cytokine and it is expected that this will result in neuroprotection.

Host(s) to be used: rodents

NIH Guidelines: II-D-4-a of the NIH Guidelines for the administration of AAVs and shRNA to rodents.

Biosafety Level and Any Additional Requirements: BSL-1 for the handling of AAVs expressing shRNAs; ABSL-1 for the intracerebroventricular administration of AAV to rodents with subsequent ABSL-1 housing.

Concerns or Discussion: None

Training: All training completed.

Approval: All approved as recommended (Yes-10, No-0, Abstain-0)

Principal Investigator: [REDACTED]

Project Title: Generating Transgenic Opsin Reporter Lines in *Nematostella vectensis*

BIO number not yet assigned

Vector/Agent(s) to be used: Plasmids

Name and Function of Transcribed Nucleic Acids: inserted genes, promoters, fluorescent tag (mOrange2), Poly(A) tail; opsin promoter/fluorescent tag.

Host(s) to be used: DH-5alpha chemically competent *E. coli*, *N. vectensis*

NIH Guidelines:

- III-D-4-a for the introduction of recombinant nucleic acids into sea anemone, *N. vectensis*.
- III-F-8 Appendix C-II for maintenance of plasmids in *E. coli* K12 strains

Biosafety Level and Any Additional Requirements: BSL-1 containment and practices.

Concerns or Discussion: None

Training: All training completed.

Approval: All approved as recommended (Yes-10, No-0, Abstain-0)

Principal Investigator: [REDACTED] BIO5687 Renewal

Project Title: Induction of T-Cell Tolerance to an auto-antigen by AAV hepatic gene therapy

BIO5687

Vector/Agent(s) to be used: Adeno-associated virus (AAV)

Name and Function of Transcribed Nucleic Acids:

Name	Function
alpha-Gliadin	prolamin, celiac disease epitope
Aquaporin 4 (AQP4)	water channel protein
Gamma Gliadin	prolamin, celiac disease epitope
Green Fluorescent Protein (GFP)	Marker protein
Interleukin-10	anti-inflammatory
Myelin Basic Protein (MBP) variant 3[4Y]	Neuro protein
Myelin Oligodendrocyte Glycoprotein (MOG)	Neuro protein

Myelin Basic Protein (MBP), Proteolipid protein isoform (PLP) and Myelin Oligodendrocyte Glycoprotein (MOG)	neuroproteins
Omega-Gliadin	prolamin, celiac disease epitope
Ovalbumin (OVA)	glycoprotein
Proteolipid protein (PLP)	Neuro protein
proteolipid protein isoform (PLP) and Myelin Oligodendrocyte Glycoprotein (MOG)	neuroprotein
truncated proteolipid protein isoform (PLP)	neuroprotein
β -galactosidase	intracellular enzyme that cleaves the disaccharide lactose into glucose and galactose

Host(s) to be used: *E. Coli* DH5 α , rodent, HEK239

NIH Guidelines:

- III-D-3-e for the generation of AAVs in the presence of a helper system
- III-D-4-a for the administration of AAV vectors into rodents.
- III-F-8 appendix C-II for plasmid maintenance in K12 strains of *E. coli*

Biosafety Level and Any Additional Requirements:

- BSL-1 for the handling of AAV vector
- BSL-2 containment and practices for handling HEK293 cells.
- ABSL-1+ for injections of AAV into rodents (Additional precaution includes use of safe engineered sharps and wiping of the injection site).
- ABSL-1i for housing following injections of AAV until after the first cage change at least 72h post-administration to allow for autoclaving of bedding, after which they will be housed at ABSL-1.
- Initial and annual bloodborne pathogen training is required.

Concerns or Discussion: More detail needed for vaccine for staff. BSL level needs to be changed for housing.

Training: All training completed.

Approval: All approved as recommended (Yes-10, No-0, Abstain-0)

Principal Investigator: David Norman RD-3522 Transcription

Project Title: Functional characterization of genes conferring virulence at low temperature

BIO number not yet assigned

Vector/Agent(s) to be used: Plasmids

Name and Function of Transcribed Nucleic Acids:

PilQ (Locus Tag Rsp597_012730)	type IV pilus secretin. Part of the pil operon that produces the Type IV pili for twitching movement. Involved in virulence
Catalase Kat G (Locus tag Rsp673_12405)	Catalase-peroxidase (HPI). It has catalase and peroxidase activity. It is crucial for defense against oxidative bursts,
pilQ, PilP, PilO, PilN and PilM genes	type IV secretion core. Core operon that produces the Type IV pili for twitching movement. Involved in virulence
TssC (RSP673_21190)	Tssc is one of the conserved core components of the Type VI Secretion System (T6SS) and functions as part of the structural apparatus. T6SS is involved in intraspecific competition and virulence of <i>Ralstonia solanacearum</i>
T6SS genes tssE, tssF and tssG	Core genes of the type 6 secretion system
RipY	putative type 3 effector involved in host range of GMI1000

Host(s) to be used: *Ralstonia solanacearum*, P597, GMI1000

NIH Guidelines: III-E-2-b-(3) for work in plants with recombinant modified non-exotic microorganisms; III-F-8, Appendix C-II for recombinant DNA work in K12 strains of *E. coli*.

Biosafety Level and Any Additional Requirements: PBSL-2 containment and practices.

Concerns or Discussion: Suggested adding III-F-8, Appendix C-II from the NIH Guidelines. Discussion of select agents.

Training: All training completed.

Approval: Voted on by UF Orlando IBC. Community members are not required but considered best practice. All approved as recommended (Yes-6, No-0, Abstain-0)

Principal Investigator: Piyush Jain BIO5522 Renewal

Project Title: Detection of nucleic acids using CRISPR

BIO5522

Vector/Agent(s) to be used: Plasmids

Name and Function of Transcribed Nucleic Acids: Nucleocapsid gene, Encodes the nucleocapsid of Coronavirus/ SARS-CoV-1 Coronavirus/ MERS-CoV Coronavirus.

Host(s) to be used: N/A

NIH Guidelines: The project is exempt from the NIH Guidelines per section III-F-2 (Experiments in which recombinant or synthetic nucleic acids have not been modified or manipulated to enable penetration of cellular membranes)

Biosafety Level and Any Additional Requirements: BSL-2+ for work with RG3 HIV samples; BSL-2 for work with RG2 coronavirus-derived infectious agents and HCV samples and RNA; BSL-1 for work with RG1 biological reagents (gamma-irradiated and heat inactivated Coronavirus 2, inactivated Zeptomatrix panels, non-genomic virus RNA, quantitative PCR (qPCR) Control RNA from inactivated SARS Coronavirus, and CoV DNA control plasmids) (the + designation necessitates the use of mucous membrane protection, the use of safe engineered sharps, and the substitution of glassware with plastic). Initial and annual bloodborne pathogens training is required

Concerns or Discussion: BSL was discussed.

Training: All training completed.

Approval: All approved as recommended (Yes-10, No-0, Abstain-0)

Principal Investigator: [REDACTED] BIO5642 Renewal

Project Title: Engineering immune cells for cancer immunotherapy

BIO number not yet assigned**Vector/Agent(s) to be used:** Lentivirus, second-generation (Gag/Pol/Rev expressed in a single helper plasmid)**Name and Function of Transcribed Nucleic Acids:**

28bbz-2A-EGFP	Modifications to a signaling protein important in T cell receptor activation
28bbz-EGFP	Modifications to a signaling protein important in T cell receptor activation
28z-2A-EGFP	Modifications to a signaling protein important in T cell receptor activation
28z-EGFP	Modifications to a signaling protein important in T cell receptor activation
EGFP	Protein that gets excited at 395nm wavelength of light and releases green fluorescent signal
m28bbz-2A-EGFP	Modifications to a signaling protein important in T cell receptor activation
m28bbz-EGFP	Modifications to a signaling protein important in T cell receptor activation

Host(s) to be used: Primary human cells, Jurkat T cell line and 293T cells for packaging. Primary murine splenocytes and T cells, and 293T cells for packaging.**NIH Guidelines:** III-E-1 for the transduction of cells with 2nd generation lentiviral vectors; III-D-3-a for the generation of Lentiviral vectors (2nd gen) in the presence of a helper system; III-D-4-a for the use of recombinant materials in rodents; III-F-8 Appendix C-I for maintenance of LVV-modified cell lines. The pMD2.G plasmid encoding the VSV-G gene is subject to export control regulations.**Biosafety Level and Any Additional Requirements:** BSL-2 for experiments involving primary human cells and established human cell lines, BSL-2+ for *in vitro* experiments involving the formation and use of the lentiviral vectors with additional precautions to include mucous membrane protection and substitution of glassware with plastic ware, and use of safe engineered sharps; ABSL-2+ for *in vivo* procedures, including intravenous administration of LVV transduced cells in rodents (additional precautions include use of safe engineered sharps and wiping of injection sites) with ABSL-2 housing afterward. ****NOTE**** If negative RCL data is provided to the IBC, containment may be reduced to ABSL-1+ for *in vivo* inoculations (use of safe engineered sharps and wiping of injection site) ABSL-1 for subsequent housing rodents receiving murine cells transduced

with RCV negative LVV and ABSL-1+Hu for rodents receiving human cells transduced with RCV negative LVV. Initial and annual bloodborne pathogens training is required.

Concerns or Discussion: None

Training: All training completed.

Approval: All approved as recommended (Yes-12, No-0, Abstain-0)

Principal Investigator: Michael Bubb

Project Title: A Phase 1 Study of FT819 in B-cell Mediated Autoimmune Diseases

BIO number not yet assigned

Vector/Agent(s) to be used: genetically-modified cells

Name and Function of Transcribed Nucleic Acids: anti-CD19 CAR at the TRAC locus, simultaneous knockout of the endogenous TCR and controlled CAR expression under the endogenous TCR alpha promoter

Host(s) to be used: human patients

NIH Guidelines: III-C-1 for the administration of a genetically modified human cell line to human research participants.

Biosafety Level and Any Additional Requirements: BSL-2 for the administration of a genetically modified cell therapy to human research participants (safe sharps should be used). Initial and annual Bloodborne Pathogens training is required.

Concerns or Discussion: None

Training: All training completed.

Approval: All approved as recommended (Yes-12, No-0, Abstain-0)

Principal Investigator: Karina Alvina BIO5764 Renewal

Project Title: Neuromodulation of smell and reward

Approval: Pulled from Agenda**Amendments****Principal Investigator:** [REDACTED] – BIO5876**Title:** Post-transcriptional regulation of gene expression in neuromuscular disease**BIO5876**

Summary: New AAV's. BSL-1 for *in vitro* studies with purchased AAVs and plasmids, as well as for *in vitro* studies with murine cell lines; BSL-2 for *in vitro* studies involving human cell lines; BSL-2+ for generation and *in vitro* use of 2nd gen lentiviral vectors and studies involving human cell lines transduced with 2nd gen lentivirus; ABSL-1+ for injections of rodents with AAVs, with subsequent ABSL-1i housing (after 72 hours the bedding must be changed and inactivated, at which point ABSL-1 housing is appropriate). The applicable extra (+) precautions include the use of mucous membrane protection, safe sharps and wiping of the injection site. The following sections of the NIH Guidelines apply to this project: III-D-3-a for packaging lentivirus in the presence of a host-vector system; III-D-4-a for the *in vivo* application of AAVs in rodents; III-E-1 for the transduction of cell lines with lentivirus and the use of CRISPR to modify cell lines; III-F-8, Appendix C-I for the maintenance of plasmids in cell culture; III-F-8, Appendix C-II for the use of plasmid vectors with nonpathogenic K-12 lineage *E. coli*.

Concerns or Discussion: None**Training:** All training completed.**Approval:** All approved as recommended (Yes-11, No-0, Abstain-0)**Principal Investigator:** Frank White – BIO6122**Title:** Research on Clavibacter species, wilt pathogens of maize, tomato, alfalfa and potato**BIO6122**

Summary: Updates Personnel and permit and adds recombinant work. This amended project is approved at BSL-2/PBSL-2 containment and practices and emphasize the collection and inactivation of run-off water from plants inoculated with genetically-modified strains. This amended

project is approved at BSL-2/PBSL-2+ containment and practices. Emphasis placed on the collection and inactivation of run-off water from plants inoculated with genetically-modified strains. This work falls under the following sections of the NIH Guidelines: III-E-2-b-(3), for plants associated with recombinant or synthetic nucleic acid molecule-modified non-exotic microorganisms that have a recognized potential for serious detrimental impact on managed or natural ecosystems; III-F-6 and III-F-8 appendix C-II for plasmid cloning and maintenance using nonpathogenic K-12 lineage *E. coli*. The work must be performed in full accordance with conditions stipulated in the accompanying permit.

Concerns or Discussion: Discussed if BSC is certified.

Training: All training completed.

Approval: Conditional approval pending confirmation of BSC certification (Yes-11, No-0, Abstain-0)

Principal Investigator: Janice Zale – BIO6099

Title: Mature Citrus Transformation and Gene Editing Service

BIO6099

Summary: Vectors added. This amended project is approved at BSL-1/PBSL-1 containment and practices for the work involving the transfer of recombinant nucleic acid molecules to plants and subsequent propagation of genetically modified plants. The following section of the NIH Guidelines apply to this project: III-E-2-a is for delivery of recombinant/synthetic nucleic acids in plants and plant-associated microorganisms.

Concerns or Discussion: None

Training: All training completed.

Approval: Voted on by UF Orlando IBC. Community members are not required but considered best practice. All approved as recommended (Yes-6, No-0, Abstain-0)

Principal Investigator: Jorge Lascano – BIO7138

Title: A Phase 1 Study of Inhaled KB4808 for the Treatment of Alpha-1 Antitrypsin Deficiency

Approval: Pulled from Agenda

Principal Investigator: [REDACTED] – BIO6425

Title: The role of glucan phosphatases in metabolism

BIO6425

Summary: Personnel changes, addition of plasmids. This amended project is approved at the following biosafety containment and practices: BSL-1 for CRISPR work, work with K-12 *E. coli* strains, and AAV transductions with murine cell lines; BSL-2 for all work with human cell lines including transduction, transfection, and CRISPR work, and *in vitro* work with adenoviral vectors; ABSL-2 for injection of unmodified human cells into rodents with subsequent housing at ABSL-1+Hu; ABSL-1+ for the injection of AAV in rodents with subsequent ABSL-1 for housing after non-systemic injection (Intracerebroventricular) and subsequent ABSL-1i for housing after systemic injection (Intravenous, Subcutaneous, Intramuscular, Intranasal, with inactivation of bedding materials for the first 72 hours post-inoculation for rodents, and disinfection of caging (the '+' necessitates the use of mucus membrane protection, use of safe sharps, and substitution of glassware with plastic whenever possible); ABSL-2+ for injection of adenovirus and LVV into rodents and with ABSL-2 for subsequent housing for injection of 2nd gen LVV and ABSL-1 for subsequent housing for injection of 3rd gen LVV. This work falls under the following sections of the NIH Guidelines: III-D-1-a for the use of recombinant adenoviral vectors; III-D-3-a for packaging lentiviral and adenoviral vectors in the presence of a host-vector system; III-D-3-e for packaging AAVs in the presence of a host-vector system; III-D-4-a for the administration of recombinant lentiviral and recombinant AAVs to rodents; III-D-4-b for the administration adenoviral vectors to rodents; III-E-1 for the transduction of lentiviral and rAAV vectors in HEK293 cells; Application of genome editing in eukaryotic hosts (such as CRISPR/Cas9. technologies); III-F-8, C-I for transfection of plasmids into cell lines and maintenance of genetically modified cell lines *in vitro*; III-F-8, C-II for the addition of cloning and maintenance of plasmids in K-12 strain *E. coli* (Top10). Personnel must complete annual Bloodborne Pathogens training.

Concerns or Discussion: Would like clarification of lentivirus use.

Training: All training completed.

Approval: Conditional approval pending the lab address comments in Question 15. (Yes-11, No-0, Abstain-0)

Principal Investigator: [REDACTED] – BIO6129

Title: Use of rodent models of disease to develop biomarkers and therapeutics

BIO6129

Summary: Personnel changes. Addition of lentivirus, and updates to models and AAVs. This amended project is approved at the following biosafety containment and practices: BSL-1 for *in vitro* experiments involving the use of AAV and CRISPR; BSL-2 for handling and *in vitro* use of human cell lines; BSL-2+ for the *in vitro* handling of 2nd generation lentiviral vectors and subsequently transduced cells (the extra "+" precautions emphasize the substitution of glassware with plasticware, the use of mucous membrane protection, and the use of safe engineered sharps); ABSL-1+ for the systemic injection of AAV into rodents, and for the transfer of transgenic material between rodents, with subsequent ABSL-1i housing (bedding to be collected and inactivated for 72 hours post administration, as well as cage disinfection; after the 72 hour period, ABSL-1 housing is appropriate); ABSL-1+Hu for procedures transferring human cell lines to rodents; ABSL-2+ for the systemic injection of second-generation lentiviral vectors transduced cells into rodents with ABSL-2 for subsequent housing (the extra + precautions emphasize the substitution of glassware with plasticware, the use of mucous membrane protection, the use of safe engineered sharps, and wiping of injection sites). If negative replication competent lentivirus (RCV) data are provided to the IBC, *in vivo* inoculations of transduced cells may be reduced to ABSL-2+/ABSL-1 for procedures and housing, respectively (solely applicable to the use of LVV transduced murine cells in animals). This work falls under the following sections of the NIH Guidelines: III-D-3-a for packaging LVV in the presence of a host vector system; III-D-4-a for the use of AAV in rodents, as well as the adoptive transfer of tissues from transgenic donor rodents to recipient rodents; III-D-4-b for the use of LVV transduced cells in rodents; III-E-1 for the transduction of human cell lines with lentivirus as well as *in vitro* CRISPR; III-F-8 Appendix C-I for the *in vitro* maintenance of genetically modified cells. Annual bloodborne pathogens training is required.

Concerns or Discussion: None

Training: All training completed.

Approval: Conditional approval pending confirmation of BSC certification (Yes-11, No-0, Abstain-0)

The meeting adjourned at 2:12pm